

Incidence of Lactobacilli in Buffalo Milk Cheddar Cheese

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ABSTRACT

The incidence of Lactobacilli in buffalo milk cheddar cheese was studied. The counts of Lactobacilli varied from 3×10^6 to 50×10^6 /g of cheese. The number of Lactobacilli was highest during the first and the fourth months of ripening in batch I and II respectively; subsequently started to decline. In a total of 105 isolates *L. casei*, *L. plantarum*, *L. leichmanii*, *L. buchneri*, *L. fermenti*, *L. brevis* and *L. cellobiosus* were 58, 37, 1, 5, 1, 1 and 2 respectively. Proteolytic abilities varied significantly with regard to strains, types of milk and the period of incubation. *L. buchneri*-8 exhibited highest proteolysis in cow milk and *L. casei*-3 in buffalo milk. The pH of the cheeses varied between 5.2 to 5.6.

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INTRODUCTION

Lactobacilli, the most dominant group of bacteria in mature cheddar cheese, when occurring naturally in cheese would be supposed to be able to better survive cheese as an environment and selected strains might grow profusely in cheese if used as an additive. Such strains, if possess desirable characteristics, can be successfully employed to hasten the ripening process in cheese, Johns and Cole (1959), Feagan and Dawson (1959), Lane and Hammer (1935), Sherwood (1939) and Jagannath *et al.* (1973).

The present study was planned to examine the buffalo milk Cheddar cheese samples for the number and types of Lactobacilli present in it and examine their proteolytic ability in cow and buffalo milk, which would serve as the basis for selecting more proteolytic strains as starter additives to hasten the rate of ripening in the cheddar cheese

made from buffalo milk probably with some enhancement in organoleptic score.

Published data on the types of Lactobacilli occurring in buffalo milk cheddar cheese and their characteristics are not available.

MATERIALS AND METHODS

Collection, handling and treatment of cheese samples:

Fourteen buffalo milk cheddar cheese samples of varying age were collected from two batches of cheese manufactured at the Amul Dairy, Anand. The samples were immediately analysed using the procedure described in the Standard Methods for the Examination of Dairy Products (1960).

Counting, isolation and identification of lactobacilli:

The counting was done using the medium of Mann, Rogosa and Sharpe (MRS) (1960).

A few colonies from each plate were randomly picked up and replated on the

MRS Agar for purification and then transferred into MRS Agar slabs incubated at 30° C for 5 days and then stored at -8° C till further use for identification.

All the isolates were identified to species level using the scheme of Sharpe *et al.* (1968).

Judging proteolytic ability of the selected isolates:

From among the isolates two strains of each species were examined for proteolytic abilities in cow and buffalo skim milk at 14° C for a period of seventyfive days. Sixteen hour old milk cultures of the isolates were inoculated into autoclaved skim milk at the rate of 1% (v/v). Proteolysis was judged at 15 days interval using the method of Hull (1947). Colour intensity was measured on Bausch and Lomb Spectronic-20 Spectrophotometer at 650 nm.

The pH of the cheese was measured by the method of Franklin and Sharpe (1963).

Statistical analysis:

The statistical analysis was done by the methods of Snedecor and Cochran (1967).

RESULTS AND DISCUSSION

The data presented in the above table indicate that the pH of cheese in both the batches ranged between 5.2 to 5.6 during the ripening period of seven months, as normally observed in cow milk cheese.

The count of Lactobacilli were quite high in numbers, almost comparable to the other two counts viz. of total viable bacteria and, the Enterococci which ranged between 7.3×10^6 to 1.2×10^8 /g and 1×10^4 to 8.8×10^5 /g respectively. In the cheese from batch

TABLE 1

Lactobacillus count of Buffalo Milk Cheese at Different Stages of Ripening. (incubated at 30° C for 5 days).

Age of cheese Months	pH of cheese		Lactobacillus Count/g ($\times 10^6$)	
	Batch I	Batch II	Batch I	Batch II
1	5.2	5.2	50	5
2	5.4	5.4	35	28
3	5.4	5.5	18	27
4	5.5	5.5	23	29
5	5.5	5.5	3	12
6	5.5	5.5	3	6
7	5.6	5.6	3	4

I, they were highest in numbers i.e. 50×10^6 /g at one month and then gradually declined till seventh month. The high number present could probably be ascribed to the higher rate of contamination with these bacteria during the manufacturing process of cheese, especially from vats, handlers and other sources on the plant. Batch II had little lower count in the beginning, then increasing upto fourth month and subsequently showing a decline probably due to both, accumulation of toxic metabolites and depletion of nutrients.

Table 2 contains the data on identification of the Lactobacilli. The isolates were identified to species level using the scheme of Sharpe *et al.* (1968). The results observed indicated that ability to produce NH_3 from arginine varied with the strains. Homo—as well as Heterofermentative strains showed this ability. Sharpe (1962) accepted some exceptions in using this test as a substitute for the type of lactic fermentation. About 50% of the *L. plantarum* strains failed to ferment arabinose.

TABLE 2

Identification of lactobacillus isolates from buffalo milk cheddar cheese.

Number of isolates out of 105	Growth at		Fermentation of					NH ₃ from	Asculin Hydro- lysis	Lactic	Species
	0.4%	15°C	45°C	Treha- lose	Sucrose	Xylose	Meli- biose	Lactose	Arabi- nose		
58	—	+	—(36) + (22)	—(47) + (11)	—(19) + (39)	—(57) + (1)	—	—(36) + (22)	—(56) + (2)	—(1) + (57)	<i>Lactobacillus casei</i>
37	—(21) + (16)	+	—(20) + (17)	—(17) + (20)	+	+	+	—(2) + (35)	—(18) + (19)	+	<i>Lactobacillus plantarum</i>
5	—(1) + (5)	+	+	—	—(2) + (3)	—	+	—(1) + (4)	+	+	<i>Lactobacillus buchneri</i>
1	—	—	+	+	+	+	—	+	—	+	<i>Lactobacillus leichmannii</i>
1	+	—	+	—	+	—	—	+	—	—	<i>Lactobacillus fermenti</i>
1	+	+	—	—	+	+	—	+	+	+	<i>Lactobacillus brevis</i>
2	+	+	—	+	+	+	+	+	—	+	<i>Lactobacillus cellobiosus</i>

Figures in the parentheses show number of isolates.

Hm = Homofermentative.

Ht = Heterofermentative.

The results exhibited in Table 2 indicated that *L. casei* and *L. plantarum* were the most dominant species encountered. Fiftyeight *L. casei* and thirty-seven *L. plantarum* made a total of 95 out of 105 total isolates. *L. leichmanii*, *L. buchneri*, *L. fermenti*, *L. brevis* and

L. cellobiosus were 1, 5, 1, 1 and 2 respectively.

Table 3 covers the data on proteolytic abilities of selected strains from among the isolates. *L. buchneri*-8 exhibited highest proteolytic ability in cow milk and *L. casei*-3 in buffalo milk.

TABLE 3

Proteolytic activity of selected *Lactobacillus* Isolates at 14°C.

Organism	Type of Milk	Time of incubation (days)				
		15	30	45	60	75
		(mg of tyrosine/5 ml TCA filtrate)				
<i>L. casei</i> -1*	Cow	0.66	0.9	1.1	1.6	2.0
	buffalo	0.64	0.8	1.0	1.8	1.8
<i>L. casei</i> -3	cow	0.23	0.5	0.7	1.1	1.5
	buffalo	0.18	0.4	0.8	2.1	2.4
<i>L. plantarum</i> -4	cow	0.24	0.61	0.72	1.2	1.9
	buffalo	0.2	0.3	0.6	0.8	1.2
<i>L. plantarum</i> -5	cow	0.15	0.25	0.42	0.82	1.3
	buffalo	0.16	0.35	0.63	0.9	1.5
<i>L. buchneri</i> -8	cow	0.32	0.72	0.93	1.4	2.4
	buffalo	0.26	0.50	0.7	0.9	1.6
<i>L. buchneri</i> -29	cow	0.45	0.89	1.2	1.6	1.8
	buffalo	0.51	0.9	1.3	1.8	2.1
<i>L. fermenti</i> -57	cow	0.7	1.1	1.5	1.8	2.6
	buffalo	0.5	0.7	0.9	1.3	1.6
<i>L. leichmanii</i> -46	cow	0.02	0.04	0.04	0.08	0.1
	buffalo	0.01	0.02	0.06	0.06	0.08
<i>L. cellobiosus</i> -96	cow	0.31	0.41	0.62	0.87	1.2
	buffalo	0.4	0.6	0.82	0.93	1.5
<i>L. brevis</i> -81	cow	0.2	0.4	0.42	0.5	0.8
	buffalo	0.3	0.3	0.5	0.58	0.9

*Number suffixed to the strain of bacteria represents isolate number.

There was a wide variation in the proteolytic abilities of different species and of different strains within a species. The release of tyrosine increased progressively with the time of incubation in all cases. The type of milk i.e. cow or buffalo also had effect on this activity.

The results presented in Table-4 showed that the strains of *Lactobacilli* significantly (5% level) differed in their average proteolytic activity. *L. casei*-1, *L. buchneri*-29 and *L. fermenti*-57 topped the list for their proteolytic activity. However, in view of the heterofermen.

Table 4

Effect of types of milk on average proteolytic activity of different strains of Lactobacilli.

Strains	mg of tyrosine/5ml TCA filtrate		Mean
	Cow milk	Buffalo milk	
<i>L. casei</i> -1	1.25	1.20	1.23
<i>L. casei</i> -3	0.80	1.17	0.98
<i>L. plantarum</i> -4	0.93	0.62	0.78
<i>L. plantarum</i> -5	0.58	0.70	0.64
<i>L. buchneri</i> -8	1.15	0.79	0.97
<i>L. buchneri</i> -29	1.18	1.32	1.25
<i>L. fermenti</i> -57	1.54	1.00	1.27
<i>L. leichmanii</i> -46	0.05	0.04	0.04
<i>L. cello-</i> <i>biosus</i> -96	0.68	0.85	0.77
<i>L. brevis</i> -81	0.46	0.51	0.48
Mean	00.862	0.820	0.841

Note: The milk samples were incubated at 14° C for 75 days.

S.E. Strain-0.0559 C.D. 0.1604

S.E. Milk x Strains-0.0790 C.D. 0.227

C.V.% = 20.91%

tative nature of the latter two strains it is suggested that *L. casei*-1 has the potentiality to be used as a proteolytic starter additive in cheddar cheese-making. Further, the said strain behaved almost uniformly in both the types of milk used.

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REFERENCES

- Feagan, J. T. and Dawson, D. J. 1959. Bacteriology of Cheddar Cheese. Some observations on the microflora during maturing. *Australian J. Dairy Technol.*, **14**: 59.
- Franklin, J. G. and Sharpe, M. E. 1963. The incidence of bacteria in cheese milk and cheddar cheese and their association with flavour. *J. Dairy Res.*, **30**: 87.
- Hull, M. E. 1947. Studies on milk proteins. II. Colorimetric determination of the partial hydrolysis of the proteins of milk. *J. Dairy Sci.*, **30**: 881.
- Jagannath, E. G., Rao Keshavmurthy, N. and Ananthramiah, S. N. 1973. Shortening the time required for preparing cheddar cheese. *Indian J. Anim. Sci.*, **43**: 306.
- Johns, S. K. and Cole, Shirley E. 1959. Lactobacilli in cheddar cheese. *J. Dairy Res.*, **26**: 157.
- Lane, C. B. and Hammer, B. W. as cited by Marth E. H. 1963. Microbiological and chemical aspects of cheddar cheese ripening: A review. *J. Dairy Sci.*, **46**: 869.
- Man, J. C. de, Rogosa, M., and Sharpe, M. E. 1960. A medium for cultivation of Lactobacilli. *J. Appl. Bact.* **23**: 130.
- Sharpe, M. E. 1962. Taxonomy of the Lactobacilli. *Dairy Sci. Abstr.*, **24**: 109.
- Sharpe, M. E., Fryer, T. F. and Smith, D. C. 1968. Identification methods for microbiologists, Part A, Academic Press Inc., London, Paegs 65-77.
- Sherwood, I. R. 1939. Lactic acid bacteria in relation to cheese flavour-II. Observations on the inoculation of the milk employed in cheese manufacture with Lactobacilli. *J. Dairy Res.*, **10**: 449.
- Standard methods for the examination of dairy products, 11th ed., APHA, 1960.
- Snedecor, G. W. and Cochran, W. G. 1967. Statistical Methods, 6th ed. Oxford and IBH Publishing Co., p. 424.